

INDUCTION OF DRUG METABOLISING CAPACITY IN THE RAT BY DI-(2-ETHYLHEXYL)PHTHALATE

Diane E. Matthew, J.B. Houston, Department of Pharmacy, University of Manchester, Manchester M13 9PL.

Numerous chemicals, including certain drugs, when administered chronically cause liver hypertrophy, proliferation of endoplasmic reticulum and induction of the cytochrome P-450 drug metabolising system. These effects have well documented consequences *in vivo* resulting in an enhanced drug metabolising capacity. Di-(2-ethylhexyl) phthalate (DEHP) is a plasticizer which readily leaches out from plastic materials and hence is ingested in significant amounts as a food contaminant and is delivered parenterally to patients on infusion therapy (Thomas et al 1978). DEHP displays many of the classic induction characteristics in the *in vitro* systems and it is believed to selectively induce formation of a specific cytochrome component (P-452) concerned primarily with fatty acid oxidation (Lake et al 1984). The consequences of DEHP on drug metabolising capacity *in vivo* are unclear and hence we have investigated the effects of multiple dosing of DEHP on the pharmacokinetics of antipyrine, a well established marker of drug metabolising capacity (Rhodes & Houston, 1983).

Male Sprague-Dawley rats (n=18) received (N-methyl-¹⁴C)-antipyrine (25mg/kg; 10 μ Ci/kg; i.p.) and were housed in all-glass metabolism cages which allowed the continuous collection of ¹⁴CO₂ (derived from N-demethylation) and urine. The half-life of antipyrine was estimated from the ¹⁴CO₂ exhalation rate (CER)-time profile. Animals were divided into 4 groups and each rat tested before, and at varying times up to 15 days while receiving daily oral doses of corn oil (5ml/kg) alone or together with DEHP (0.5, 1 or 2g/kg). On the day after the 15th dose the rats were killed, livers removed for preparation of microsomes by standard differential centrifugation.

The data shown in table 1 demonstrates that at each of the doses studied there is liver enlargement and an increase in two microsomal markers of drug metabolising capacity - cytochrome P450 content and ethoxycoumarin O-deethylase. In contrast the antipyrine CER half-life is affected only at the highest dose. The maximal reduction in half-life following DEHP dosing, which is considerably less than that observed with other classic inducers (Rhodes & Houston 1983) is maximal at 7 days and maintained until day 15. It would appear that cytochrome P-452 and any other cytochromes inducible by DEHP do not show a strong affinity for antipyrine as a substrate. Although additional studies employing other drug substrates are required, the present investigation would indicate limited consequences of DEHP induction on *in vivo* metabolism.

Table 1 Effects of various doses of DEHP on drug metabolising capacity

Dose (g/kg)	CER half-life(min)		Liver weight (% of body weight)	Cytochrome P-450 (nmoles/mg protein)	Ethoxycoumarin O-deethylase (nmoles/min/mg protein)
	Day 0	Day 15			
0	125 \pm 7	126 \pm 11	3.4 \pm 0.2	0.345 \pm 0.074	0.657 \pm 0.131
0.5	112 \pm 10	112 \pm 8	4.5 \pm 0.2 ^a	0.463 \pm 0.095 ^b	0.881 \pm 0.129 ^b
1	117 \pm 7	110 \pm 9	5.4 \pm 0.2 ^a	0.396 \pm 0.154	0.825 \pm 0.195
2	128 \pm 26	101 \pm 17 ^c	5.7 \pm 0.4 ^a	0.496 \pm 0.042 ^a	1.076 \pm 0.105 ^a

Mean \pm s.d. Statistical significance from control p<0.01(a) and p<0.05 (b) by one way ANOVA and p<0.05 (c) by two way ANOVA.

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